



Cite this: *Med. Chem. Commun.*, 2017, 8, 1332

A structurally guided dissection-then-evolution strategy for ligand optimization of smoothened receptor†‡

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Received 28th February 2017,
Accepted 20th April 2017

DOI: 10.1039/c7md00104e

rsc.li/medchemcomm

We present herein a novel dissection-then-evolution strategy for ligand optimization. Using the co-crystal structure of the smoothened receptor (SMO) as a guide, we studied the modular contribution of LY2940680 by systematically “silencing” the specific interaction between the individual residue(s) and the fragment in the ligand. Following evolution by focusing on the benzoyl part finally yielded an improved ligand **21**.

The smoothened receptor (SMO) is a transmembrane protein categorized as a member of Frizzled G protein-coupled receptors (GPCRs). SMO plays an essential role in the regulation of the evolutionarily conserved and physiologically important Hedgehog (Hh) signaling pathway.^{1,2} Disruption of this pathway results in disorders of embryonic development and tissue regeneration that are related to birth defects or cancers.^{3,4} Thus, SMO has been a well-pursued and now validated target for anticancer agent development.^{5–8} Many SMO ligands have been discovered, including the steroidal alkaloid cyclopamine and other synthetic compounds.^{9–11} Among these, vismodegib (GDC-0449)¹² and sonidegib (LDE225)¹³ have been approved by the FDA in 2012 and 2015, respectively, for the treatment of basal-cell carcinoma (BCC). Nevertheless, their clinical use has been associated with many severe side reactions,^{14,15} as well as several instances of mutation-related drug

resistance.^{16–18} In addition, many fundamental questions remain regarding understanding SMO's involvement in those physiological mechanisms.¹⁹ Development of a persistent and reliable ligand that would address these issues is highly sought after.

The evolution of SMO ligands can now be guided by the high resolution structures that were obtained recently.²⁰ As part of the achievements from the intense and flourishing research on SMO, several atomic resolution structures of SMO, including those of the transmembrane domain (TMD)^{21,22} and extracellular domain (ECD),^{23,24} and the very recent multi-domain structures have emerged.²⁵ These structures clearly defined the molecular contacts of co-crystallized SMO ligands and provided great insights into the design of tools and probes for further biophysical studies. For example, originating from the first co-crystallized SMO ligand LY2940680,^{21,26} our groups designed a super-stabilizing tool ligand TC114 (Fig. S1, ESI†) by the introduction of a new functional group. This modification established an extra interaction that restricts motion to hold the receptor's conformation and facilitate crystallization for structural studies of multi-domain SMO.²⁷ This success encouraged us to look for an additional improved ligand with potential applications in pharmacological development and probe design.

Using known SMO structures as a guide, we present herein a dissection-then-evolution strategy for ligand optimization. To this end, we first carried out a comprehensive study on LY2940680, whose interactions between its receptor residues and ligand fragments were revealed at high resolution (Fig. 1A).²¹ According to the key interactions as indicated, the molecule was dissected into four variable parts (I–IV) and one linker (Fig. 1B). Sequential “silencing”, namely introducing a chemical mutation into LY2940680 to weaken

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† The authors declare no competing interests.

‡ Electronic supplementary information (ESI) available: Materials, synthetic protocols, computational process and additional data. See DOI: 10.1039/c7md00104e



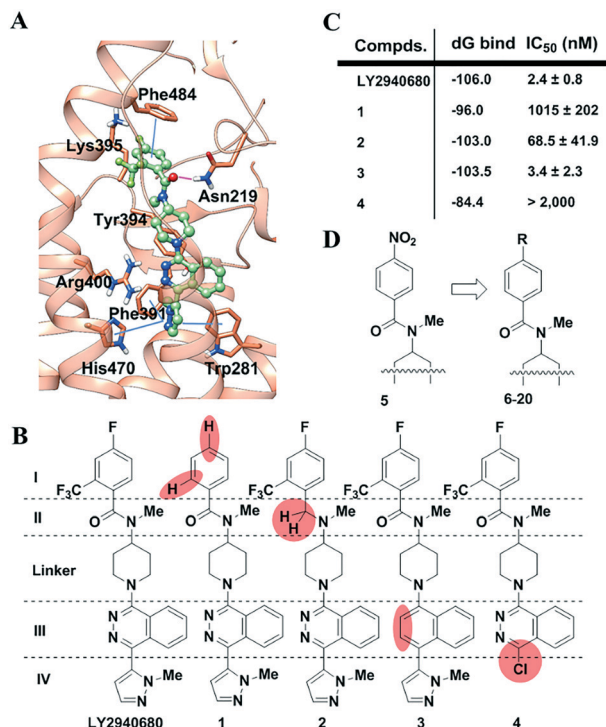


Fig. 1 Study of the SMO ligand LY2940680. (A). The key interactions observed in the co-crystal structure of SMO with LY2940680. (B). Design of LY2940680 analogs to analyze the contribution from each part. LY2940680 was dissected into four parts (I–IV) and one linker. Analogs were designed by partially “silencing” the interaction by the removal or substitution of the functional groups, as highlighted in coral in every compound. (C). The relative binding energy was calculated and the potency was evaluated by the Gli assay. IC₅₀ values represent the mean ± SEM of at least three separate experiments carried out in duplicate. All compounds were evaluated following 24 h incubation. (D). New analogs were designed by focusing on the variations in the benzoyl unit.

or eliminate a specific ligand–receptor interaction, generated a series of compounds (1–4, Fig. 1B). All of the new compounds were synthesized (Scheme S1, ESI†) and then tested for activity inhibiting the Hedgehog signaling pathway by a luciferase reporter assay using light II cells, which were NIH-3T3 cells stably transfected with a Gli-responsive firefly luciferase reporter. As clearly indicated (Fig. 1C), compounds with substituents removed on benzoyl (1) or deletion of the pyrazole (4) almost completely abolished the receptor's activity. Meanwhile, the replacement of phthalazine with naphthalene (3) only accounts for a slight reduction of the activity. This may indicate that there is little difference between the electronic interaction of Arg400 with phthalazine and the cation– π interaction with naphthalene. Surprisingly, the replacement of the amide by a tertiary amine in 2 maintained moderate activity with an IC₅₀ of about 70 nM, possibly because of compensation resulting from the missing H-bonding between the carbonyl and Asn219. Additionally, to exclude the potential interruption from the metabolism of these compounds, we checked their stability in cell media (Fig. S3, ESI†) which indicated that this group of compounds was sta-

ble and thus the dissecting effect is only dependent on the structural variation.

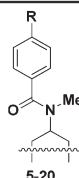
Glide XP docking followed by Prime MM-GBSA was used to estimate the relative binding affinities of the compounds mentioned above.²⁸ As shown (Fig. 1C and Table S1, ESI†), modification of pyridazine leads to the least loss of relative binding energy (ddG = 2.5 kcal mol⁻¹). In comparison, the alteration of other fragments would result in a large loss of the binding energy. For compound 4, the loss of pyrazole leads to a huge loss of ddG (21.6 kcal mol⁻¹), which is mainly from van der Waals energy (ddG VDW = 11.0 kcal mol⁻¹). Similarly, removal of substituents in benzoyl also results in a large loss of ddG (~10 kcal mol⁻¹). Interestingly, elimination of the carbonyl renders compound 2 with a unique basic tertiary amine which, under physiological conditions, produces a protonated 2-H⁺ which can still maintain a similar binding free energy to that of the parent compound LY2940680 because the solvation energy (ddG solv = -28.1 kcal mol⁻¹) compensates for the loss of coulomb energy (ddG Coulomb = 28.6 kcal mol⁻¹) of 2-H⁺.

Overall, in this context, the computational performance showed good agreement with the IC₅₀ values from cell-based functional assays. These results confirmed our hypothesis that modification in part I or IV would be valuable, while that in part II or III would be less sensitive. Furthermore, considering the space limitations of the deeply buried pocket that interacts with part IV, the more outward situated part I fragment seems to be in a better position for modification. It may not be a coincidence that our previous development of TC114 (Fig. S1, ESI†) in which a nitro group was introduced to the benzoyl yielded successful co-crystallization. However, an aromatic nitro group, like in TC114, was generally avoided by medicinal chemists since its reductive metabolite causes undesired toxicity. Thus, our aim here is to search for a better and more drug-like surrogate for the nitro group (Fig. 1D). To simplify the synthesis and characterization, all the compounds were initially designed without the 6-trifluoromethyl group on the benzoic fragment. For this reason, all the compounds were then compared to that described in ref. 5, a 4-NO₂ compound derived from TC114 by removal of the trifluoromethyl group.

A series of compounds (5–20) were then synthesized (Scheme S2, ESI†). Following our established procedure in search for stabilizing molecules on GPCRs, the compounds were first screened *via* a thermal stability assay on purified SMO (Table 1 and Fig. S2, ESI†). This assay shows the fluorescence (365/460 nm) increase during the thermal denaturation upon reaction of exposed cysteines with a thiol-specific fluorochrome, CPM dye (*N*-[4-(7-diethylamino-4-methyl-3-coumarinyl)phenyl]maleimide). The thermal stabilization capability of the test compounds was denoted as the midpoint temperature (*T*_m) at which 50% of the fluorescence increase was observed. The CPM assay has been widely used in the high throughput screening of tool compounds on membrane proteins.^{29,30}

As indicated in Table 1, almost all the compounds were able to stabilize SMO to some extent when compared to the



Table 1 Exploration of structure activity relationships (SARs) on potential analogs


Compds.	R	CPM assay T_m (°C)	MM-GBSA dG bind (kcal mol ⁻¹)	Gli-Luc reporter IC ₅₀ ^a (nM)
APO	NA	56.3	NA	NA
5	NO ₂	61.0	-100.6	235.6 ± 88.5
6	F	59.1	-93.6	1575 ± 506
7	NH ₂	58.2	-92.3	2018 ± 556
8	NHCH ₂ CO ₂ H	56.8	-88.8	>10 000
9	CHO	59.0	-97.9	2891 ± 407
10	NMe ₂	58.5	-96.3	1415 ± 338
11	SO ₂ Me	56.3	-87.3	>10 000
12	SO ₃ H	59.4	-86.6	2682 ± 898
13	SO ₂ NH ₂	57.1	-98.1	3242 ± 712
14	CO ₂ Me	60.3	-100.6	>10 000
15	CO ₂ H	56.8	-89.5	>10 000
16	CONH ₂	59.7	-97.5	3018 ± 223
17	B(OH) ₂	59.4	-101.1	870 ± 358
18	CH ₂ CHO	60.2	-99.0	>10 000
19	OC ₆ H ₅	59.6	-111.8	269.7 ± 59.1
20	OCH ₂ C ₆ H ₅	58.4	-113.2	1799 ± 443

^a IC₅₀ values represent the mean ± SEM of at least three independent experiments carried out in duplicate. All analogues were evaluated following 24 h incubation.

APO sample (T_m = 56.3 °C). Among them, 5 remained the best stabilizer (T_m = 61.0 °C). Surprisingly, little improvement was observed on the negatively charged compounds 8, 12 and 15 which, according to the design with different acidity and alternative linker length, are expected to form an ionic pair with the positively charged ϵ -amine of Lys395. This may be due to the high desolvation penalty of the highly solvated ligand anion that weakens the contribution from the ionic pair. Simple substituents, like fluorine (6), showed a slight benefit on stabilization (T_m = 59.1 °C). Other neutral compounds without a negatively induced dipole, such as 7 and 10, exhibited even less favorable interactions (T_m \approx 58 °C), while compounds with possible negative dipoles to mimic the nitro group, like sulfone 11 and ester 14, could indeed promote the expected interaction. The corresponding sulfonamide 13 and amide 16, however, showed less capability to stabilize. This may be due to an attenuated π - π interaction between the ligand and Phe484 by a relatively weaker electron-withdrawing group than the nitro in compound 5. An aldehyde is proposed to form a reversible covalent Schiff base with an amine. However, 9 did not significantly enhance the stability of SMO. 18 with an extended linker is a better ligand in this regard as it may have a higher tendency to form a covalent bond. Boronic acids, though not often, have been found in marketed drugs. Functionally, phenyl boronic acid usually has a pK_a of 9–10 that makes it protonated under physiological conditions. It

has been reported to behave as the bioisostere of phenol,³¹ and it was proposed to form a hydrogen bond or a boron–nitrogen bond at times. Thus, it could be an alternative to explore the interaction with Lys395. However, compound 17 did not provide encouraging results (T_m = 59.4 °C). Finally, two phenyl-substituted compounds 19 and 20 exhibited moderately enhanced stabilization on SMO.

To further elucidate the structure activity relationships (SARs), we performed MM-GBSA calculations and measured IC₅₀s *via* a Gli-luciferase reporter assay. As indicated (Table 1 and Fig. S6, ESI[†]), the relative binding free energy and T_m values correlate well, in a linear manner with only a few exceptions. Compounds that deviated from the correlation include the highly acidic phenyl sulfonic acid 12 and phenyl attached 19 and 20. For the latter two compounds, calculations provided an extremely low binding free energy (dG bind < -110). The major difference in the formula came from VDW and lipophilicity (Table S1, ESI[†]) when compared to those with nitro embedded 5. The low T_m value reflected that the loading of such a large rigid and hydrophobic group is not favorable for stabilizing SMO in micelles. Nevertheless, the correlations indicated that the calculation method can be a rough alternative for the prediction of a ligand's stabilizing effect.

On the other hand, the Gli-luciferase reporter assay showed a slight consistency with the CPM assay or MM-GBSA calculation. Roughly, compounds with lower binding energy behave relatively well with a few exceptions. 14 has a high T_m value and a low binding energy but exhibited no activity in the Gli assay. This may be ascribed to its *in vivo* hydrolysis to acid 15 which, in general in this assay, lost activity like 8. Overall, the nitro compound 5 remained the best with an IC₅₀ of 235 nM. Excitingly, 19 ranked second with a close IC₅₀ value, though not as potent as reflected in the MM-GBSA calculation. Other molecules were generally much less effective, in the micro molar range of IC₅₀ values in this assay.

We then further modified 19 to 21 by adding a trifluoromethyl group on the benzoyl unit (Fig. 2A). As observed in the transformation of 5 and TC114,²⁷ this evolution dramatically enhanced the activity in the Gli assay. Trifluoromethyl groups play multiple roles in pharmaceutical development. This strong electron-withdrawing group likely enhanced the π - π interaction with Phe484. In comparison with TC114, 21 could be a promising drug candidate for further evolution since 21 maintained comparable potency when the nitro group was replaced with a pharmacologically safer group. We carefully determined the docking of compound 21 in our recent full-length structure of SMO (Fig. 2B).²⁷ This compound retained all the important interactions with SMO as in the TC114 co-crystallized structure. In particular, 21 establishes two additional key interactions, both involved with the appended benzene ring. One is the cation- π interaction between Lys395 and the benzene,³² and the other is the π - π interaction with Phe484. By comparison, such interactions were not observed in other compounds. For example, a closely resembling compound S1 (Fig. S4, ESI[†]), modified



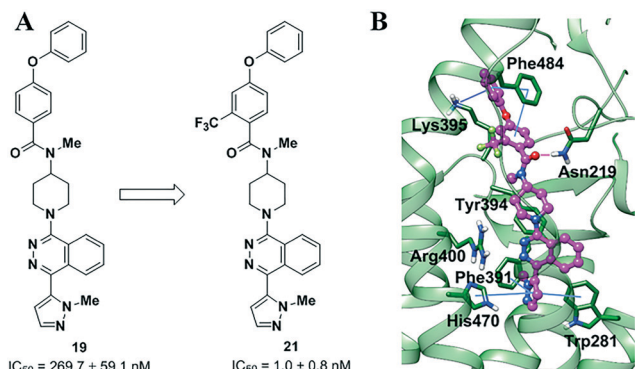


Fig. 2 Compound **21** exhibited enhanced potency. (A). Evolution of **19** by the addition of a trifluoromethyl group on the benzoyl unit generated compound **21** with IC_{50} of 1.0 nM. (B). Docking study of **21** in the two-domain SMO showed that all the original key interactions were retained. Besides, the embedded benzene ring at the outer end of a molecule of **21** established two additional interactions with Lys395 and Phe484, respectively.

from **20** by one more carbon extension in the linker, showed a dramatic decrease in efficacy in the Gli luciferase assay. In general, these extra interactions may promote ligand potency on SMO and more importantly, hold promise in applications against other disease-related SMO mutations. It should also be noted that the newly introduced benzene ring added an excellent knob for the derivation of probes for other biophysical studies. In fact, in our attempts to design tool compounds in our on-going projects, we found that derivatives based on other analogs, as shown in Table 1, would have to pay a price with a decreasing efficacy.

In conclusion, a more drug-like and better probe-oriented ligand **21** was developed for SMO *via* a structurally guided dissection-then-evolution strategy. We confirmed that the introduction of a phenyl ether into LY2940680 can strengthen its binding and stabilize the SMO. This combined computation and chemistry strategy could be employed on other scaffolds or against other targets for future projects, including imaging, structural exploration and drug discovery.

The authors (FX, RCS, SZ, and HT) are thankful to Shanghai Municipal Government, ShanghaiTech University for financial support, the Shanghai Pujiang Talent Program (16PJ1407300; SZ) and the Natural Science Foundation of Shanghai (16ZR1448500; SZ). We thank Y. Zhao for providing the NIH3T3 cell line for compound evaluation assays. We thank J. Yang and W. Tan for performing the cytotoxicity assay. We also thank Z. J. Liu, G. Song and J. Cheng for their helpful discussion and assistance in manuscript preparation.

Notes and references

- 1 Y. Zhao, C. Tong and J. Jiang, *Nature*, 2007, **450**, 252.
- 2 P. W. Ingham and A. P. McMahon, *Genes Dev.*, 2001, **15**, 3059.
- 3 M. P. di Magliano and M. Hebrok, *Nat. Rev. Cancer*, 2003, **3**, 903.
- 4 J. Jiang and C.-C. Hui, *Dev. Cell*, 2008, **15**, 801.
- 5 E. Dolgin, *Nat. Med.*, 2011, **17**, 523.
- 6 C. A. Maschiot, J. R. Pace and M. K. Hadden, *Curr. Med. Chem.*, 2015, **22**, 4033.
- 7 G. Liu, J. Yang, J. Wang, X. Liu, W. Huang, J. Li, W. Tan and A. Zhang, *J. Med. Chem.*, 2016, **59**, 11050.
- 8 H. J. Sharpe, W. Wang, R. N. Hannoush and F. J. de Sauvage, *Nat. Chem. Biol.*, 2015, **11**, 246.
- 9 J. K. Chen, J. Taipale, M. K. Cooper and P. A. Beachy, *Genes Dev.*, 2002, **16**, 2743.
- 10 J. K. Chen, J. Taipale, K. E. Young, T. Maiti and P. A. Beachy, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**, 14071.
- 11 H. Tao, Q. Jin, D.-I. Koo, X. Liao, N. P. Englund, Y. Wang, A. Ramamurthy, P. G. Schultz, M. Dorsch, J. Kelleher and X. Wu, *Chem. Biol.*, 2011, **18**, 432.
- 12 A. Dlugosz, S. Agrawal and P. Kirkpatrick, *Nat. Rev. Drug Discovery*, 2012, **11**, 437.
- 13 S. Pan, X. Wu, J. Jiang, W. Gao, Y. Wan, D. Cheng, D. Han, J. Liu, N. P. Englund, Y. Wang, S. Peukert, K. Miller-Moslin, J. Yuan, R. Guo, M. Matsumoto, A. Vattay, Y. Jiang, J. Tsao, F. Sun, A. C. Pferdekamper, S. Dodd, T. Tuntland, W. Maniara, J. F. Kelleher, III, Y.-m. Yao, M. Warmuth, J. Williams and M. Dorsch, *ACS Med. Chem. Lett.*, 2010, **1**, 130.
- 14 S. E. Gould, J. A. Low, J. C. Marsters, Jr., K. Robarge, L. L. Rubin, F. J. de Sauvage, D. P. Sutherlin, H. Wong and R. L. Yauch, *Expert Opin. Drug Discovery*, 2014, **9**, 969.
- 15 E. Pak and R. A. Segal, *Dev. Cell*, 2016, **38**, 333.
- 16 G. J. P. Dijkgraaf, B. Alicke, L. Weinmann, T. Januario, K. West, Z. Modrusan, D. Burdick, R. Goldsmith, K. Robarge, D. Sutherlin, S. J. Scales, S. E. Gould, R. L. Yauch and F. J. de Sauvage, *Cancer Res.*, 2011, **71**, 435.
- 17 H. J. Sharpe, G. Pau, G. J. Dijkgraaf, N. Basset-Seguine, Z. Modrusan, T. Januario, V. Tsui, A. B. Durham, A. A. Dlugosz, P. M. Haverly, R. Bourgon, J. Y. Tang, K. Y. Sarin, L. Dirix, D. C. Fisher, C. M. Rudin, H. Sofen, M. R. Migden, R. L. Yauch and F. J. de Sauvage, *Cancer Cell*, 2015, **27**, 327.
- 18 S. X. Atwood, K. Y. Sarin, R. J. Whitson, J. R. Li, G. Kim, M. Rezaee, M. S. Ally, J. Kim, C. Yao, A. L. S. Chang, A. E. Oro and J. Y. Tang, *Cancer Cell*, 2015, **27**, 342.
- 19 A. M. Arensdorf, S. Marada and S. K. Ogden, *Trends Pharmacol. Sci.*, 2016, **37**, 62.
- 20 J. Shonberg, R. C. Kling, P. Gmeiner and S. Loeber, *Bioorg. Med. Chem.*, 2015, **23**, 3880.
- 21 C. Wang, H. Wu, V. Katritch, G. W. Han, X.-P. Huang, W. Liu, F. Y. Siu, B. L. Roth, V. Cherezov and R. C. Stevens, *Nature*, 2013, **497**, 338.
- 22 C. Wang, H. Wu, T. Evron, E. Vardy, G. W. Han, X.-P. Huang, S. J. Hufeisen, T. J. Mangano, D. J. Urban, V. Katritch, V. Cherezov, M. G. Caron, B. L. Roth and R. C. Stevens, *Nat. Commun.*, 2014, **5**, 4355.
- 23 R. Rana, C. E. Carroll, H.-J. Lee, J. Bao, S. Marada, C. R. R. Grace, C. D. Guibao, S. K. Ogden and J. J. Zheng, *Nat. Commun.*, 2013, **4**, 3965.
- 24 P. Huang, D. Nedelcu, M. Watanabe, C. Jao, Y. Kim, J. Liu and A. Salic, *Cell*, 2016, **166**, 1176.



- 25 E. F. X. Byrne, R. Sircar, P. S. Miller, G. Hedger, G. Luchetti, S. Nachtergaele, M. D. Tully, L. Mydock-McGrane, D. F. Covey, R. P. Rambo, M. S. P. Sansom, S. Newstead, R. Rohatgi and C. Siebold, *Nature*, 2016, 535, 517.
- 26 M. H. Bender, P. A. Hippskind, A. R. Capen, M. Cockman, K. M. Credille, H. Gao, J. A. Bastian, J. M. Clay, K. L. Lobb, D. J. Sall, M. L. Thompson, T. Wilson, G. N. Wishart and B. K. R. Patel, *Cancer Res.*, 2011, 71, 2819.
- 27 X. Zhang, F. Zhao, Y. Wu, J. Yang, G. W. Han, S. Zhao, A. Ishchenko, L. Ye, X. Lin, K. Ding, V. Dharmarajan, P. R. Griffin, C. Gati, G. Nelson, M. S. Hunter, M. A. Hanson, V. Cherezov, R. C. Stevens, W. Tan, H. Tao and F. Xu, *Nat. Commun.*, 2017, DOI: 10.1038/ncomms15383.
- 28 J. Li, R. Abel, K. Zhu, Y. Cao, S. Zhao and R. A. Friesner, *Proteins: Struct., Funct., Bioinf.*, 2011, 79, 2794.
- 29 X. Zhang, R. C. Stevens and F. Xu, *Trends Biochem. Sci.*, 2015, 40, 79.
- 30 A. I. Alexandrov, M. Mileni, E. Y. T. Chien, M. A. Hanson and R. C. Stevens, *Structure*, 2008, 16, 351.
- 31 Y. Kong, J. Grembecka, M. C. Edler, E. Hamel, S. L. Mooberry, M. Sabat, J. Rieger and M. L. Brown, *Chem. Biol.*, 2005, 12, 1007.
- 32 J. P. Gallivan and D. A. Dougherty, *Proc. Natl. Acad. Sci. U. S. A.*, 1999, 96, 9459.

